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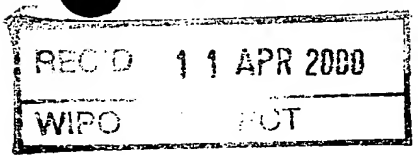

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I, KAY WARD, TEAM LEADER EXAMINATION SUPPORT AND SALES hereby certify that annexed is a true copy of the Provisional specification in connection with Application No. PP 9463 for a patent by JOHNSON & JOHNSON RESEARCH PTY LIMITED filed on 26 March 1999.

WITNESS my hand this
Third day of April 2000

K Ward

KAY WARD
TEAM LEADER EXAMINATION
SUPPORT AND SALES

AUSTRALIA

PATENTS ACT 1990

PROVISIONAL SPECIFICATION

FOR THE INVENTION ENTITLED:-

"CODEINONE REDUCTASE FROM ALKALOID POPPY"

The invention is described in the following statement:-

Technical Field

The present invention relates to codeinone reductase from alkaloid poppy plants, the polynucleotides encoding the enzyme and to production of alkaloids from transformed poppy plants.

Background

The search for useful drugs of defined structure from plants began with the isolation of morphine from dried latex, or opium, of the opium poppy *Papaver somniferum* in 1806 (Sertürner). The narcotic analgesic morphine and the antitussive and narcotic analgesic codeine, the antitussive and apoptosis inducer noscapine (Ye et al., 1998), and the vasodilator papaverine are currently the most important physiologically active alkaloids from opium poppy. Of these four alkaloids, only papaverine is prepared by total chemical synthesis for commercial purposes. Opium poppy, therefore, serves as one of the most important renewable resources for pharmaceutical alkaloids. Per annum, 90-95% of the approximately 160 tons of morphine that are purified are chemically methylated to codeine, which is then used either directly or is further converted to a variety of derivatives such as dihydrocodeinone and 14-hydroxydihydrocodeinone that find use as antitussives and analgesics (Kutchan, 1998). The illicit production of morphine for acetylation to heroin is unfortunately almost ten times that amount, more than 1200 tons per year (Zenk, 1994).

The enzymatic synthesis of morphine in opium poppy has been almost completely elucidated by M.H. Zenk and coworkers and is summarized by Kutchan (1998). Opium poppy produces more than 100 different alkaloids that are derived from

the amino acid L-tyrosine and have the tetrahydrobenzylisoquinoline alkaloid, (S)-reticuline, as a common intermediate. There are three NADPH-dependent reductases involved in the conversion of (S)-reticuline to morphine. (S)-Reticuline must first be converted to (R)-reticuline before the phenanthrene ring with the correct stereochemistry at C-13 can be formed. The inversion of stereochemistry at C-1 of (S)-reticuline occurs by oxidation to the 1,2-dehydroreticulinium ion followed by stereospecific reduction to the R-epimer by 1,2-dehydroreticulinium ion reductase [EC 1.5.1.27] (De-Eknamkul and Zenk, 1992). The second reduction occurs after formation of the phenanthrine nucleus with stereospecific reduction of salutaridine to salutaridinol by salutaridine reductase [EC 1.1.1.248] (Gerardy and Zenk, 1993). The third reduction is the penultimate step in the biosynthetic pathway to morphine, the reduction of codeinone to codeine by codeinone reductase [EC 1.1.1.2471] (Figure 1; Lenz and Zenk, 1995a,b). The substrate for codeinone reductase, codeinone, exists in an equilibrium with its positional isomer neopinone. *In vitro,* as codeinone is reduced, this equilibrium is continually driven from neopinone towards codeinone until the substrates are depleted (Gollwitzer et al., 1993). Each of the known enzymes of morphine biosynthesis has been detected in both *P. somniferum* plants and cell suspension culture, yet plant cell cultures have never been shown to accumulate morphine (Kutchan, 1998). Sequences of genes encoding cytochrome P450 reductases have been published in PCT/AU98/000705 which is hereby incorporated by reference.

To date, no other genes specific to morphine biosynthesis in opium poppy have been isolated. Tyrosine/dopa decarboxylase has been investigated at the molecular genetic level, but is involved in multiple biochemical processes in this plant (Facchini

and De Luca, 1994). Morphine, along with the chemotherapeutic agents vincristine, vinblastine and camptothecin, is one of the most important alkaloids commercially isolated from medicinal plants. Isolation of the genes of morphine biosynthesis would facilitate metabolic engineering of opium poppy to produce plants with specific patterns of alkaloids and could ultimately lead to an understanding of the inability of plant cell cultures to accumulate morphine.

It is an object of the present invention to overcome or ameliorate at least one of the disadvantages of the prior art, or to provide a useful alternative.

Summary of the Invention

The narcotic analgesic morphine is the major alkaloid of the opium poppy *Papaver somniferum*. Its biosynthetic precursor codeine is currently the most widely used and effective antitussive agent. Along the morphine biosynthetic pathway in opium poppy, codeinone reductase catalyzes the NADPH-dependent reduction of codeinone to codeine. At least 10 codeinone reductase alleles are present in the genome of the poppy *Papaver somniferum*. Isolation, characterization and functional expression of four of the 10 genes encoding codeinone reductase as described herewith enables methods for controlling alkaloid production in opium poppy plants and cultures by providing a target for genetic manipulation.

Thus, according to a first aspect, there is provided an isolated and purified polynucleotide or a variant, fragment or analog thereof, encoding a codeinone reductase enzyme from an alkaloid poppy plant.

The polynucleotide may be selected from the group consisting of genomic DNA (gDNA), cDNA, or synthetic DNA. Preferred polynucleotides are selected from (a) the

polynucleotide sequences shown in FIGS: 10 to 15; (b) the polynucleotide sequences which hybridize under stringent conditions to the complementary sequences of (a); and (c) polynucleotide sequences which are degenerate to polynucleotide sequences of (a) or (b). It will be understood however that the sequences may be expressed in the absence of the native leader sequences or any of the 5' or 3' untranslated regions of the polynucleotide. Such regions of the polynucleotide may be replaced with exogenous control/regulatory sequences in order to optimise/enhance expression of the sequence in an expression system.

The preferred alkaloid-producing poppy plant is *Papaver somniferum.*

It will also be understood that analogues and variants of the polynucleotide encoding a codeinone reductase from alkaloid poppy plants fall within the scope of the present invention. Such variants will still encode an enzyme with codeinone reductase properties and may include codon substitutions or modifications which do not alter the amino acid encoded by the codon but which enable efficient expression of the polynucleotide encoding codeinone reductase enzyme in a chosen expression system. Other variants may be naturally occurring, for example allelic variants or isoforms.

According to a second aspect there is provided an isolated and purified polynucleotide, or a variant, analog or fragment thereof, which codes for prokaryotic or eukaryotic expression of a codeinone reductase enzyme from an alkaloid poppy plant, wherein the polynucleotide is expressed in an environment selected from the group consisting of the extracellular environment, an intracellular membranous compartment, intracellular cytoplasmic compartment or combinations thereof.

The polynucleotide encoding a codeinone reductase may be coupled to another nucleotide sequence which would assist in directing the expression of the reductase with respect to a particular cellular compartment or the extracellular environment.

According to a third aspect there is provided an isolated and purified polynucleotide which is complementary to all or part of the sequence of a polypeptide according to the first aspect.

Such complementary polynucleotides are useful in the present invention as probes and primers, as antisense agents or may be used in the design of other suppressive agents such as ribozymes and the like.

According to a fourth aspect there is provided a recombinant DNA construct comprising the polynucleotide according to any one of the first to third aspects.

Preferably the recombinant DNA construct is a viral or plasmid vector. Such a vector may direct prokaryotic or eukaryotic expression of the polynucleotide encoding a codeinone reductase or it may prevent or reduce its expression. The vector may also be selected from pCAL-c, pGEM-T or pFastBac1. Preferably the promoter used to control expression of the codeinone reductase gene is selected from nos, cauliflower mosaic virus or subterranean clover mosaic virus.

According to a fifth aspect there is provided an isolated and purified codeinone reductase enzyme, being a product of prokaryotic or eukaryotic expression of the polynucleotide of any one of first to third aspects or a DNA construct of the fourth aspect.

The codeinone reductase may be expressed in and by a variety of eukaryotic and prokaryotic cells and organisms, including bacteria, yeasts, insect cells, mammalian and

other vertebrate cells, or plant cells. Preferably the expression system is a plant expression system and even more preferred is an alkaloid poppy plant. A suitable alkaloid poppy plant is *Papaver somniferum*.

Variants of the codeinone reductase enzyme which incorporate amino acid deletions, substitutions, additions or combinations thereof, are also contemplated. The variants can be advantageously prepared by introducing appropriate codon mutations, deletions, insertions or combinations thereof, into the polynucleotide encoding the codeinone reductase enzyme. Such variants will retain the properties of the codeinone reductase enzyme, either *in vivo* or *in vitro*, and may have improved properties. Other variants may be naturally occurring, for example allelic variants or isoforms.

For expression of codeinone reductase activity, a fragment of the polynucleotide encoding a codeinone reductase may be employed, such fragment encoding functionally relevant regions, motifs or domains of the reductase protein. Similarly, fragments of the codeinone reductase enzyme resulting from the recombinant expression of the polynucleotide may be used. Functionally important domains of codeinone reductase may be represented by individual exons or may be identified as being highly conserved regions of the protein molecule. Those parts of the codeinone reductase which are not highly conserved may have important functional properties in a particular expression system.

According to a sixth aspect there is provided a cell transformed or transfected with a polynucleotide according to any one of the first to third aspects or a DNA construct according to the fourth aspect.

Cells which may be transfected or transformed with a polynucleotide encoding a codeinone reductase are bacterial, yeast, animal or plant cells. For preference the cells are plant cells. Even more preferred are cells from an alkaloid poppy plant, such as *Papaver somniferum*.

According to the seventh aspect, there is provided a callus transformed or transfected with a polynucleotide according to any one of the first to third aspects or a DNA construct according to the fourth aspect.

According to the eighth aspect, there is provided a plant transformed or transfected with a polynucleotide according to any one of the first or third aspects or a DNA construct according to the fourth aspect wherein the plant exhibits altered expression of the codeinone reductase enzyme. For preference, the altered expression manifests itself in overexpression of the codeinone reductase enzyme. However, reduced expression of codeinone reductase can also be achieved if the plant is transformed or transfected with a polynucleotide which is complementary to the polynucleotide encoding the reductase.

Even more preferably, the transformed or transfected plant is an alkaloid poppy plant, wherein the plant has a higher or different alkaloid content when compared to a plant which has not been so transformed or transfected.

Preferably the transformed or transfected plants having higher or different alkaloid content are *Papaver somniferum*.

According to the ninth aspect, there is provided a method for preparing plants which overexpress a codeinone reductase enzyme, comprising transfecting or transforming a plant cell, a plant part or a plant, with the polynucleotide according to any

one of the first to third aspects or a DNA construct according to the fourth aspect.

Preferably the plant overexpressing codeinone reductase is an alkaloid poppy plant and most preferably the poppy plant is *Papaver somniferum*. Suitable promoters to control the expression of the codeinone reductase gene may be derived from for example nos, cauliflower mosaic virus or subterranean clover mosaic virus. Other virus promoters may also be suitable. Further, the use of the endogenous promoter may also be appropriate in certain circumstances. Such a promoter may be co-isolated with the gDNA encoding the codeinone reductase enzyme.

According to the tenth aspect, there is provided a method of altering the yield or type of alkaloid in a plant comprising transforming or transfecting a plant cell, a plant part or a plant with a polynucleotide, or a variant, analog or fragment thereof, encoding a codeinone reductase enzyme, or with a polynucleotide which binds under stringent conditions to the polynucleotide encoding the enzyme.

According to the eleventh aspect, there is provided a method of increasing the yield of alkaloid in a plant comprising transforming or transfecting a plant cell, a plant part or a plant with a polynucleotide, or a variant, analog or fragment thereof, encoding a codeinone reductase enzyme wherein the enzyme is overexpressed in the plant.

According to the twelfth aspect, there is provided a method of altering type or blend of alkaloid in a plant comprising transforming or transfecting a plant cell, a plant part or a plant with a polynucleotide or a variant, analog or fragment thereof, encoding a codeinone reductase enzyme or with a polynucleotide which binds under stringent conditions to the polynucleotide encoding said enzyme.

According to the thirteenth aspect, there is provided a stand of stably reproducing alkaloid poppies transformed or transfected with a polynucleotide according to any one of the first to third aspects or a DNA construct according to the fourth aspect, having altered expression of the codeinone reductase enzyme.

According to the fourteenth aspect, there is provided a stand of stably reproducing alkaloid poppies transformed or transfected with a polynucleotide according to any one of the first to third aspects or a DNA construct according to the fourth aspect, having a higher or different alkaloid content when compared to a plant which has not been so transformed or transfected.

Preferably the stably reproducing alkaloid poppy is *Papaver somniferum*.

According to the fifteenth aspect, there is provided straw of stably reproducing poppies according to the fourteenth aspect having a higher or different alkaloid content when compared to the straw obtained from an alkaloid poppy which has not been transformed or transfected.

According to the sixteenth aspect, there is provided a concentrate of straw according to the fifteenth aspect having a higher or different alkaloid content when compared to the concentrate of straw obtained from an alkaloid poppy which has not been transformed or transfected.

According to the seventeenth aspect, there is provided an alkaloid when isolated from the straw according to the fifteenth aspect or the concentrate according to the sixteenth aspect. Preferably the alkaloid is selected from the group consisting of morphine, codeine, oripavine and thebaine.

According to the eighteenth aspect, there is provided a method for production of poppy plant alkaloids comprising the steps of ;

a) harvesting capsules of an alkaloid poppy plant transformed or transfected with a polynucleotide according to any one of the first to third aspects, or a DNA construct according to the fourth aspect, to produce a straw where the poppy plant is such a plant that the straw has a higher or different alkaloid content when compared to the straw obtained from a poppy plant which has not been transformed or transfected; and

b) chemically extracting the alkaloids from the straw.

According to the nineteenth aspect, there is provided a method for the production of poppy alkaloids comprising the steps of;

a) collecting and drying the latex of the immature capsules of an alkaloid poppy plant transformed or transfected with a polynucleotide according to any one of the first to fourth aspects, to produce opium wherein the poppy plant is such a plant that the opium has a higher of different alkaloid content when compared to the opium obtained from a poppy plant which has not been transformed or transfected; and

b) chemically extracting the alkaloids from the opium.

For preference the alkaloid is morphine, codeine, oripavine or thebaine, but it will be understood that other intermediates in the alkaloid metabolic pathway are also within the scope of the present invention, as are mixtures of alkaloids.

According to a twentieth aspect, the invention provides the polynucleotide sequence encoding codeinone reductase comprised in microbial deposit No. 12737.

According to a twenty-first aspect, the invention provides the polynucleotide sequence encoding codeinone reductase comprised in microbial deposit No. 12738.

According to a twenty-second aspect, the invention provides the polynucleotide sequence encoding codeinone reductase comprised in microbial deposit No. 12739.

According to a twenty-third aspect, the invention provides the polynucleotide sequence encoding codeinone reductase comprised in microbial deposit No. 12740.

Unless the context clearly requires otherwise, throughout the description and the claims, the words 'comprise', 'comprising', and the like are to be construed in an inclusive sense as opposed to an exclusive or exhaustive sense; that is to say, in the sense of "including, but not limited to".

Brief Description of the Figures

Figure 1. Biosynthetic pathway leading from S-Reticuline to morphine in the opium poppy, *Papaver somniferum*.

The reduction of codeinone to codeine by codeinone reductase drives the non-enzymatic equilibrium between neopinone and codeinone in a physiologically forward direction. The demethylation of thebaine and codeine are each thought to be catalyzed by cytochrome P450-dependent enzymes.

Figure 2. Partial amino acid sequences of native codeinone reductase.

Codeinone reductase was purified to apparent electrophoretic homogeneity from cell suspension cultures of opium poppy and hydrolyzed with endoproteinase Lys-C. The resultant peptide mixture was resolved by HPLC and the amino acid sequences of seven peptides were obtained.

Figure 3. Amino acid sequence homology of codeinone reductase internal peptides.

Codeinone reductase peptides 3, 7, 14, 16, and 17 aligned with the reductase subunit of the 6'-deoxychalcone synthase complex from alfalfa, glycyrrhiza and soybean allowing the relative positioning of these internal peptides from opium poppy.

Figure 4. Amino acid sequence comparison of codeinone reductase isoforms.

The amino acid sequences derived from translation of the nucleotides sequences of *cor1.1-1.4* as compared to the reductase subunit of the 6'-deoxychalcone synthase complex from soybean *(6'dcs)* indicate the very high sequence identity between isoforms (95-96%) and this reductase of phenylpropanoid metabolism (53%). The complete amino acid sequence of *cor1.1* is shown, but only those non-identical residues of the four subsequent sequences.

Figure 5. Genomic DNA gel blot analysis of the codeinone reductase gene family in opium poppy.

Genomic DNA isolated from opium poppy cell suspension cultures was hybridized to *cor1.1* full-length cDNA and was visualized by phosphorimaging. The numbers following the restriction enzyme names indicate the number of recognition sites that occur within the *cor1.1* reading frame. This high stringency Southern analysis indicates the presence of at least ten alleles in the opium poppy genome.

Figure 6. RNA gel blot analysis of distribution of codeinone reductase transcript in a mature opium poppy.

The gel blot was prepared from RNA isolated from leaf mid rib, lateral root and 12 cm of stem tissue directly beneath the receptacle of an opium poppy plant 2 days after

petal fall. 50 μg of total RNA were loaded per gel lane. The RNA was hybridized to *cor1.1* full length cDNA and was visualized by phosphorimagery.

Figure 7. SDS-PAGE analysis of fractions from the purification of codeinone reductase fusion protein from *E. coli*.

Codeinone reductase was expressed as a C-terminal fusion with a 25 amino acid calmodulin-binding peptide in *E. coli* BL21 (DE3)pLysS. Protein bands were visualized with coomassie brilliant blue R-250. Lane 1, 15 μg crude protein from an extract of *E. coli* BL21 (DE3)pLysS containing the codeinone reductase cDNA before IPTG induction; lane 2, 10 μg crude protein from an extract of *E. coli* BL21 (DE3)pLysS containing the codeinone reductase cDNA 3 h after IPTG induction; lane 3, 5 μg protein from the calmodulin affinity chromatography eluate after concentration using a Centriprep 30 column (Amicon); lane 4, Rainbow Marker protein standards (Amersham). Arrow indicates position of codeinone reductase fusion protein.

Figure 8. Chemical structures of alkaloids serving as substrates for codeinone reductase.

Of the twenty-six potential substrates tested, only seven were transformed by codeinone reductase. The names of the untransformed compounds are given in the Description of Preferred Embodiments. Codeinone is the physiological substrate for this enzyme in most, if not all, varieties of opium poppy. Morphinone also serves as a physiological substrate in Tasmanian varieties. The K_m values provided for those seven substrates were determined for COR1.3.

Figure 9. Proposed alternative biosynthetic pathway leading from thebaine to morphine in opium poppies from Tasmania.

This alternative biosynthetic pathway was proposed after oripavine was discovered in Tasmanian varieties of opium poppy (Brochmann-Hanssen, 1984). Codeinone reductase from non-Tasmanian varieties can also catalyze the reduction of morphinone to morphine (Lenz and Zenk, 1995b). COR1.1-COR1.4 each catalyzed this reduction with equivalent specific activity. The demethylation of thebaine and codeine are thought to be catalyzed by cytochrome P450-dependent enzymes.

Figure 10. cDNA sequence of *cor1.1*.

Figure 11. cDNA sequence of *cor1.2*.

Figure 12. cDNA sequence of *cor1.3*.

Figure 13. cDNA sequence of *cor1.4*.

Figure 14. Partial cDNA sequence of *cor1.5*.

Figure 15. Partial cDNA sequence of *cor1.6*.

Description of the Preferred Embodiments

cDNAs that encode codeinone reductase were isolated. Four full-length reading frames and two partial clones (FIGS 10 to 15) were isolated that represent six alleles from a gene family that may have at least 10 members. An analysis of RNA and enzyme activity from various stages of developing opium poppy seedlings and roots, stem, leaf and capsule of mature poppy plants indicated that transcript from these alleles is present throughout the plant at all developmental stages, with the highest total enzyme activity being in the capsule after petal fall. This would suggest that morphine biosynthesis occurs in all major plant organs starting within the first seven days after seed germination. Biosynthesis of morphine continues throughout the life cycle of this annual with the highest biosynthetic activity taking place in the capsule after petal fall,

consistent with the amount of biosynthetic enzyme present. The amount of extractable RNA remained high in the capsule until three days after petal fall, after which time the quantity of extractable RNA decreased rapidly.

A biochemical analysis of four functionally expressed alleles, *cor1.1-cor1.4*, revealed no significant differences in the temperature or pH optima, K_m values or substrate specificity of the isoforms. All isoforms were able to reduce morphinone to morphine.

Purification and Amino Acid Sequence Analysis of Opium Poppy Codeinone Reductase

Codeinone reductase was purified to apparent electrophoretic homogeneity from opium poppy cell suspension cultures and the amino acid sequence of seven endoproteinase Lys-C-generated peptides was determined (Figure 2). A comparison of these amino acid sequences with those available in the GenBank/EMBL sequence database allowed a relative positioning of peptides 7, 14 and 16 due to sequence homology with an NADPH-dependent reductase from members of the Fabaceae - alfalfa, glycyrrhiza and soybean (6'-deoxychalcone synthase) that synthesizes 4,2',4'-trihydroxychalcone in co-action with chalcone synthase (Figure 3) (Welle et al., 1991). PCR primers were then designed based on the codeinone reductase peptide sequences. The sequences of the primers used in the first round of PCR were:

```
5'-GAA CTT TTT ATA ACT TCT AA-3'  (derived from Peptide 14) and
    G   C   C   C   G   C
                T

3'-GTG GTC TAA CGT CAI CGT TCI CCT TT-5'  (derived from Peptide 7)
    A                   A G     C
```

Resolution of an aliquot of the first PCR experiment by agarose gel electrophoresis revealed a mixture of DNA products, none of which was the expected band of approximately 480 bp. This was presumably due to the relatively low specificity of the degenerate primers coupled to a low abundance of codeinone reductase transcript. Another aliquot of the first PCR reaction mixture was, therefore, used as template for nested PCR with the following primers:

```
5'-GAA CTT TTT ATA ACT TCT AA-3'  (same as Peptide 14 primer above) and
     G   C   C   C   G   C
                 T

3'-CAI CAC TTA GTT CAC CTT TAC-5'  (nested primer derived from Peptide 16)
         G       C       C
```

to yield an approximately 360 bp DNA fragment and the following primers to yield an approximately 180 bp DNA product:

```
5-'GTI GTI AAC CAA GTI GAA ATG AGI CCI AC-3'  (nested primer derived from
              T   G       G     TC              Peptide 16) and

3'-GTG GTC TAA CGT CAI CGT TCI CCT TT-5'  (same as Peptide 7 primer above)
     A                   A G     C
```

The results from the nested PCR were bands of the expected size. The translation of the nucleotide sequences of these PCR products indicated that they encode codeinone reductase.

Isolation of cDNAs Encoding Codeinone Reductase

Screening of approximately 200,000 clones of a primary cDNA library prepared from opium poppy RNA isolated from capsule and cell suspension culture did not result in the identification of codeinone reductase clones. Likewise, difficulty was also

confronted with detecting a band on RNA gel blots that corresponds to the size expected for codeinone reductase. In order to overcome the apparent problem of low steady state levels of codeinone reductase transcript, RACE-PCR was used to generate both the 5'- and 3'-portions of the cDNA (Frohman, 1993). A series of non-degenerate primers based on the nucleotide sequence information determined for the PCR product generated as described in the previous section were used for 5'- and 3'-RACE. The nucleotide sequence of the resultant 5'- and 3'- partial clones were thus determined in three major fragments and suggested the presence of isoforms. The full length cDNA clones were then generated by RT-PCR using the following primers and RNA isolated from opium poppy cell suspension culture as template:

5'-ATG GAG AGT AAT GGT GTA CCT-3' (located at the 5'-terminus) and

3'-TCT ACC ATT CAC TCC TGA CAG-5' (located in the 3'-flanking region)

followed by nested PCR with the following primer pair:

5'-ATG GCT AGC ATG GAG AGT AAT GGT GTA CCT ATG-3' (located at the
Nhe 1 5'-terminus) and

3'-CTT CTC AAG ACC CTA CTC TTC CTA CCT AGG GAA-5' (located at the
Bam HI 3'-terminus).

The PCR product was digested with the restriction endonucleases *Nhe* I / *Bam* HI, ligated into *Nhe* I / *Bam* HI digested pCAL-c and transformed into *Escherichia coli* BL21(DE)pLysS. Each cDNA was hence constructed in frame in front of DNA encoding a 25 amino acid long calmodulin-binding peptide to facilitate eventual heterologous protein purification. Single colonies were grown in 3 ml medium and were assayed for the ability to reduce codeinone. Of forty colonies tested, ten were found to

contain functional enzyme. Nucleotide sequence determination of these ten cDNAs resulted in the identification of four alleles encoding codeinone reductase. The analogous PCR products had also been prepared with the cDNAs placed behind the calmodulin-binding peptide gene in pCAL-n-EK, but only the C-terminal fusion proteins bound the calmodulin affinity resin, indicating that the amino terminus of the fusion protein lies within the folded polypeptide.

By sequence comparison, codeinone reductase clearly belongs to the aldo/keto reductase family, a group of structurally and functionally related NADPH-dependent oxidoreductases. Members of this family possess three consensus sequences that are also positionally conserved: aldo/keto reductase consensus 1 (amino terminus) - G (F,Y)R(H,A,L)(L,I,V,M,F)D(S,T,A,G,C)(A,S) X X X X X E X X (L,I,V,M) G [*cor1.1* - G Y R H F D T A A A Y Q T E E C L G]; aldo/keto reductase consensus 2 (central) - (L,I,V,M,F,Y) X X X X X X X X X (K,R,E,Q) X (L,I,V,M) G (L,I,V,M) (S,C) N (F,Y) [*cor1. 1* - M E E C Q T L G F T R A I G V C N F]; aldo/keto reductase consensus 3 (carboxy terminus) - (L,I,V,M) (P,A,I,V) (K,R) (S,T) X X X X R X X (G,S,T,A,E,Q,K) (N,S,L) X X (L,I,V,M,F,A) [*cor1. 1* - V V K S F N E A R M K E N L K I]. This third consensus sequence is centred around a lysine residue, the modification of which has been shown to affect the catalytic efficiency of aldose and aldehyde reductases (Morjana et al., 1989).

The four functional full-length cDNAs (*cor1.1, cor1.2, cor1.3 and cor1.4*) encoding codeinone reductase share approximately 95-96% sequence identity (Figure 4). These sequences are comprised in microbial deposit Nos. DSM 12737, DSM 12738, DSM 12739 and DSM 12740 respectively, deposited at Deutsche Sammlung von

Mikroorganismen und Zellkulturen GmbH (DSMZ) of Mascheroder Weg 1b, D-38124 Braunschweig, Germany on 16 March 1999. In addition, a similar cDNA generated by PCR (*cor2*) was 70% identical to the codeinone reductase cDNAs, but was not functional. These opium poppy cDNAs were 53% identical to soybean NADPH-dependent reductase 6'-deoxychalcone synthase (Welle et al., 1991) (Figure 4), 33% identical to rat 3-hydroxysteroid dehydrogenase [EC 1.1.1.50], 38% identical to bovine prostaglandin F synthase [EC 1.1.1.188], 37% identical to apple D-sorbitol-6-phosphate dehydrogenase [EC1.1.1.200], 38% identical to bacterial *(Pseudomonas putida)* morphine 6-dehydrogenase [EC1.1.1.218] and 35% identical to yeast *(Pichia stipitis)* xylose reductase (Amore at al., 1991).

Genomic DNA Analysis and Gene Expression Pattern

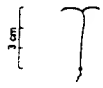
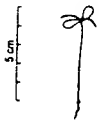
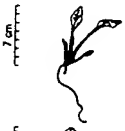
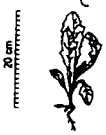
Genomic DNA was used as template for a PCR analysis of *cor1.1-cor1.4*. Each gene was found to contain one intron that was conserved in size (443 bp) and location (beginning after nucleotide +561) within the open reading frame, but not in nucleotide sequence. In comparison, *cor2* contained two introns beginning after nucleotides +321 and +514. Genomic DNA gel blot analysis using *cor1.1* as hybridization probe resulted in a complex hybridization pattern that suggests the presence of at least ten genes that could encode codeinone reductase in opium poppy (Figure 5). From the isolation and nucleotide sequence analysis of cDNA clones, it is certain that at least six of these ten genes are expressed in the plant and plant cell suspension culture. (Two additional partial cDNAs (*cor1.5* and *cor1.6*; FIGS 14 and 15) were generated by RT-PCR using plant RNA as template.) When the peptide sequences presented in Figure 2 are compared with the translations of the cDNA sequences in Figure 4, it is clear that a

mixture of isoforms was purified for amino acid sequence analysis. From the initial biochemical analysis of codeinone reductase, evidence for only two isoforms in the poppy plant and one isoform in poppy cell suspension culture was observed (Lenz and Zenk, 1995b).

RNA gel blot analysis indicated the presence of a very weakly hybridizing RNA of approximately 1.4 kb in poppy leaf, root and stem of a mature plant two days after petal fall (Figure 6). Since *cor1* transcript was apparently present at very low levels, further analysis was undertaken by nested RT-PCR. Morphinan alkaloids begin to accumulate rapidly in poppy seedlings four to seven days after germination (Rush et al., 1985; Wieczorek et al., 1986). An analysis of codeinone reductase enzyme activity and transcript accumulation showed that enzyme activity is at 310 pkat/g dry tissue weight (dwt) already at day seven after germination (Table 1). This activity remains at that level throughout a three week growth period, then decreases to 148 pkat/g dwt by the eighth week. In comparison, opium poppy cell suspension culture also contains 330 pkat/g dwt enzyme activity. Transcript was detected by RT-PCR for *cor1.1-cor1.4* at all developmental stages. Since two PCR amplifications were necessary in order to detect *cor1* transcript, a comparative quantitation was not undertaken.

The distribution of codeinone reductase enzyme activity and transcript was also investigated in mature opium poppy plants two days after petal fall. On a dry tissue weight basis, most activity was present in the capsule (730 pkat/g dwt), then the lateral root (560 pkat/g dwt) followed by stem and leaf lamina (Table 2). Again, no differences could be found in the distribution pattern of the four isoforms by RT-PCR.

Table 1. Analysis of codeinone reductase enzyme activity and transcript in developing opium poppy and in plant suspension culture.

Plant material	Plant age (days)	Specific activity (pkat/mg)	Total activity (pkat/g dwt)	Transcript detection [a]
	7	11	310	+
	14	9	330	+
	21	8	310	+
	56	12	150	+
	7	10	330	+

[a] Presence of transcript in each RNA population was determined qualitatively by performing two nested PCR amplifications as described in Material and Methods.

Table 2. Analysis of codeinone reductase enzyme activity and transcript in opium poppy two days after petal fall

Plant part	Specific activity (pkat/mg)	Total activity (pkat/g dwt)	Transcript detection [a]
 Capsule	25	730	+
Stem [b]	30	250	+
Leaf lamina	10	120	+
Lateral root	90	560	+

[a] Presence of transcript in each RNA population was determined qualitatively by performing two nested PCR amplifications as described in Material and Methods.

[b] Stem tissue beginning at the receptacle and extending 12 cm downwards was extracted. Plants were approximately 120 cm high.

Functional Characterization of the Codeinone Reductase Alleles

The four codeinone reductase isoform-calmodulin-binding peptide fusion proteins were purified from *E. coli* lysates in one step with a calmodulin affinity column. Beginning with 250 mg total protein in the bacterial extract, 10.5 mg codeinone reductase with a specific activity of 5.2 nkat/mg protein could be obtained in 73% yield. Aliquots from a typical purification analyzed by SDS-PAGE are shown in Figure 7. Codeinone reductase purified by this method is nearly homogeneous and demonstrated properties that compared favourably to those of the native enzyme (Lenz and Zenk, 1995b).

The temperature optimum, pH optimum and K_m values for codeinone, codeine, NADPH and NADP were determined for each of the isoforms (K_m values are indicated in Table 3). Significant differences in these values were not found. For all isoforms, the temperature optimum for reduction (physiologically forward reaction) was 28°C, for oxidation (physiologically reverse reaction) was 30°C, the pH optimum for reduction was 6.8 and for oxidation was 9.0. The isoforms were also tested for their ability to transform morphinan alkaloids structurally related to codeinone and codeine. The reductive reaction with NADPH as cofactor functions with morphinone, hydrocodone and hydromorphone as substrate. The oxidative reaction with NADP as cofactor functions with morphine and dihydrocodeine as substrate. The K_m values for, and structures of, these additional substrates with COR1.3 are shown in Figure 8. In all cases, the physiologically forward reaction yielded lower K_m values than the physiologically reverse reaction, with codeinone having the lowest K_m value at 48 μM. No differences in temperature or pH optimum were observed whether codeinone or

morphinone were used as substrate in the assay. NADH could not substitute for NADPH with any of the isoforms. Tritium was enzymatically transferred to codeinone from [4*R*-^{3}H]NADPH, but not from [4*S*-^{3}H]NADPH, indicating that codeinone reductase stereospecifically abstracts the pro-R hydrogen from the cofactor.

Table 3. Comparison of properties of codeinone reductase isoforms

	COR1.1	COR1.2	COR1.3	COR1.4
Amino acid identity (%)	100	95	96	96
K_m codeinone (μM)	58	62	48	50
K_m NADPH (μM)	180	220	205	197
K_m codeine (μM)	220	200	187	140
K_m NADP (μM)	53	58	45	55
Calculated M_r	35,808	35,704	35,797	35,705
Calculated pI	6.25	5.71	6.32	6.33

The reduction of codeinone to codeine is the last of three NADPH-dependent reductions that occur along the biosynthetic pathway leading from (S)-reticuline to morphine in opium poppy. The two other potential substrates for reduction, the 1,2-dehydroreticulinium ion and salutaridine (Figure 1), or for the physiologically reverse reaction, salutaridinol and (R)-reticuline, were tested as substrates; with the codeinone reductase isoforms. None of these alkaloids served as substrate indicating that codeinone reductase can catalyze only one reductive step in morphine biosynthesis. In addition, the following analogs were also inactive: (*S*) and (*R*)-norreticuline, (S)-reticuline and norcodeine.

Since codeinone reductase showed sequence similarity to several members of the aldo/keto reductase family, a series of substrates were tested to reflect members from carbohydrate and steroid metabolism. D-Sorbitol-6-phosphate, D-xylose, prostaglandin D1, 5-androstene-3β,17β-diol, 5α-androstan-17β-ol-3-one, 5α-cholestane-3β-ol, β-estradiol, cyclohexanone and 2-cyclohexene-1-one were not transformed by codeinone reductase. The highest amino acid sequence identity (53%) was, however, to the reductase subunit of the 6'-deoxychalcone synthase complex from soybean (Welle et al., 1991). In order to test for a functional evolutionary relationship between isoflavonoid and alkaloid anabolism, codeinone reductase was analyzed for the ability to substitute for the reductase in the formation of 6-deoxychalcone in co-action with either native chalcone synthase or native stilbene synthase from *Pinus sylvestris*. In the presence of 4-coumaryl-CoA, malonyl-CoA, NADPH, chalcone synthase and codeinone reductase or cinnamoyl-CoA, malonyl-CoA, NADPH, stilbene synthase and codeinone reductase, formation of product was not observed. Likewise, the reductase of the 6'-deoxychalcone synthase complex could neither reduce codeinone in the presence of NADPH nor oxidize codeine in the presence of NADP.

Example 1

Purification of Native Enzyme and Amino Acid Sequence Analysis

Cell suspension cultures of the opium poppy *Papaver somniferum* were routinely grown in either 1-litre conical flasks containing 400 ml of Linsmaier-Skoog medium (Linsmaier and Skoog, 1965) over 7 days at 23°C on a gyratory shaker (100 rpm) in diffuse light (750 lux). Differentiated opium poppy plants were grown outdoors in

Upper Bavaria. Seedlings were grown on substrate from 7 to 56 days in a greenhouse at 20°C, 65% relative humidity and 12 h cycles of light and dark.

A mixture of codeinone reductase isoforms was purified from opium poppy cell suspension cultures exactly according to Lenz and Zenk (1995b). The purified enzyme preparation was subjected to SDS/PAGE to remove traces of impurities and the coomassie brilliant blue R-250-visualized band representing codeinone reductase was digested *in situ* with endoproteinase Lys-C as reported in (Eckerskorn and Lottspeich, 1989, Dittrich and Kutchan, 1991). The peptide mixture thereby obtained was resolved by reversed phase HPLC [column, Merck Lichrospher RP18; 5µm (4 x 125 mm); solvent system, (A) 0.1% trifluoroacetic acid, (B) 0.1% trifluoroacatic: acid / 60% acetonitrile; gradient of 1 % per min; flow rate of 1ml/min] with detection at 206 nm. Microsequencing of seven of the peptides thus purified was accomplished with an Applied Biosystems model 470 gas-phase sequencer.

Example 2

Generation of Partial and Full-Length cDNAs from Opium Poppy

Partial cDNAs encoding codeinone reductases from opium poppy were produced by PCR using cDNA produced by reverse transcription of total RNA isolated from 3 to 5-day-old suspension cultured cells. DNA amplification using either *Taq* or *Pfu* polymerase was performed under the following conditions: 4 min at 94°C, 35 cycles of 94°C, 30 sec; 45°C, 30 sec; 72°C, 1 min. At the end of 35 cycles, the reaction mixtures were incubated for an additional 5 min at 72°C prior to cooling to 4°C. Reamplification of DNA using nested primers was performed as above, but the primer annealing temperature was raised from 45 to 55°C. The amplified DNA was then resolved by

agarose gel electrophoresis, the bands of approximately the correct size were isolated and subcloned into pGEM-T (Promega) prior to nucleotide sequence determination. The specific sequences of the oligodeoxynucleotide primers used are indicated above.

Total RNA was isolated and RNA gels were run and blotted as previously described (Pauli and Kutchan, 1998). Genomic DNA was isolated and DNA gels were run and blotted according to Bracher and Kutchan (1992). cDNA clones were labelled by random-primed labelling with [α-^{32}P]dCTP and oligodeoxynucleotides were end-labelled with [γ-^{32}P]ATP. Hybridized RNA on Northern blots and DNA on Southern blots were visualised with a Raytest BAS-1500 phosphorimager. The entire nucleotide sequence on both DNA strands of full-length cDNA clones in either pGEM-T or pCAL-c was determined by dideoxy cycle sequencing using internal DNA sequences for the design of deoxyoligonucleotides as sequencing primers.

The sequence information requisite to the generation of full-length cDNAs was derived from the nucleotide sequences of the partial cDNAs generated as described above. The complete nucleotide sequence of one reading frame was determined using codeinone reductase specific oligodeoxynucleotide primers in 5'- and 3'-RACE-PCR experiments with a Marathon™ cDNA amplification kit (Clontech). RACE-PCR was performed using the PCR cycles described above. The amplified DNA was then resolved by agarose gel electrophoresis and the band of the approximate expected size was isolated, subcloned into pGEM-T and sequenced.

Nested primer pairs were then used to generate full-length clones for heterologous expression by RT-PCR using opium poppy cell suspension culture RNA as template. The final primers used in clone amplification contained the restriction

endonuclease recognition sites *Nhe* I and *Bam* HI that were appropriate for subcloning directly into the pCAL-c (Stratagene) expression vector. The specific sequences of these primers are indicated above. RT-PCR was carried out using the PCR cycles given above. The amplified DNA was then resolved by agarose gel electrophoresis and the band of the correct size (972 bp) was excised and isolated for further subcloning into the expression vector.

Example 3

Heterologous Expression and Enzyme Purification

Full-length cDNAs generated by RT-PCR were ligated into p-CAL-c and transformed into the *E.coli* strain BL21(DE3)pLysS. For enzyme assays, single colonies were picked and grown in 3 ml Luria-Bertani medium containing 100 µg/ml ampicillin at 37°C to an OD_{590} of 0.8. For protein purification, single colonies were picked and grown in 1 l Luria-Bertani medium containing 100 µg/ml ampicillin at 37°C to an OD_{590} of 1.8. Cells were collected by centrifugation 5 min at 4,000 x g and 4°C. The bacterial pellet was resuspended in either 0.1 M potassium phosphate buffer pH 6.8 for the reduction of codeinone or 0.1 M glycine buffer pH 9 for the oxidation of codeine. The bacterial pellet from a 3 ml culture was resuspended in 0.5 ml buffer and that from a one litre culture in 100 ml buffer. The cells were ruptured by sonication. Cellular debris was removed by centrifugation 5 min at 4,000 x g and 4°C and the supernantant used directly for either affinity chromatography purification using the Affinity™ Protein Expression and Purification System according to the manufacturer's instructions (Stratagene) or for enzyme activity measurements according to Lenz and Zenk (1995b).

Example 4

Enzyme Assay and Product Identification

The oxidative and reductive reactions catalyzed by codeinone reductase were assayed according to Lenz and Zenk (1995b). The oxidation of codeine to codeinone by heterologously expressed enzyme in a crude bacterial extract was used for large scale production of enzymic product for structure elucidation by ^{1}H NMR, ^{13}C NMR and mass spectrometry. The enzyme assays were extracted twice with two volumes of $CHCl_3$, the combined organic phase was reduced *in vacuo* and resolved by semipreparative HPLC using the following gradient: [column, Knauer LiChrosopher 100 RP18 endcapped; 5 µm (16 x 250 mm); solvent system, (A) 97.99% (vlv) H_2O 2% CH_3CN, 0.01% (v/v) H_3PO_4, (B) 1.99% (v/v) H_2O, 98% CH_3CN, 0.01% H_3PO_4; gradient: 0-9 min 0-8% B, 9-24 min 8% B, 24-45 min 8-25% B, 45-75 min 25% B, 75-75.3 min 25-0% B, 75.3-90 min 0% B; flow 4.5 ml/min] with detection at 204 nm using authentic codeine (retention time, 38 min) and codeinone (retention time, 49 min) as reference materials. In this manner, 10 mg codeinone was enzymically produced and purified.

Codeinone-^{1}H(360MHz, $CDCl_3$) 1.87 (1H,dd $J_{15a/15e}$12.2, $J_{15e/16a}$3.1, H-15*e*), 2.08 (1H,ddd, $J_{15a/16a}$ 4.5, $J_{15a/15e}$ 12.2, H-15_a) 2.29 (1H, ddd, $J_{15a/16a}$ 12.3, $J_{15e/16a}$ 3.1, $J_{16a/16e}$ 3.1, $J_{16a/16e}$, 11.8,H-16_a), 2.35 (1H, dd, $J_{10a/10e}$ 18.5,$J_{9/10a}$ 5.9, H-10_a), 2.47 (3H, s, CH_3N-), 2.63 (1H, dd, $J_{16a/16e}$ 11.8 $J_{15a/16e}$ 4.5,H-16_e), 3.12 (1H, d, $J_{10a/10e}$ 18.5,H-10_e), 3.21 (1H, m, H-14), 3.43 (1H, m, H-9), 3.85 (3H, s, CH_3O-), 4.71 (1H, s, H-5), 6.09 (1H, dd, $J_{7/8}$ 10.1, $J_{7/14}$ 2.8, H-7), 6.62 (1H, d, $J_{1/2}$ 8.3, H-1), 6.66 (1H, dd, $J_{7/8}$ 10.1, $J_{8/14}$ 1.5, H-8), 6.68 (1H, d, $J_{1/2}$ 8.3, H-2); ^{13}C(90.6 MHz, $CDCl_3$) 20.5 (C-10), 33.8 (C-15), 41.3 (C-14), 42.8 (NMe), 43.0 (C-13), 46.8 (C-16), 56.8 (OMe), 59.1 (C-9), 88.0 (C-5), 114.8 (C-2),

119.9 (C-1), 125.7 (C-11), 128.9 (C-12), 132.6 (C-7), 142.6 (C-3), 144.9 (C-4) 148.7 (C-8), 194.4 (C-6); EI-MS (70 eV),m/z 297 (M^+, 100%, 282 (8), 268 (9), 254 (8), 238 (9), 229 (23), 214 (17), 188 (15) 165 (11), 152 (13), 139 (16), 128 (22), 115 (41).

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THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS

1. An isolated and purified polynucleotide, or a variant, fragment or analog thereof, encoding a codeinone reductase enzyme from an alkaloid poppy plant.

2. A polynucleotide according to claim 1, selected from the group consisting of genomic DNA, cDNA, or synthetic DNA.

3. A polynucleotide according to claim 1 or claim 2, selected from the group consisting of:

(a) the polynucleotide sequences shown in FIGS 10 to 15;

(b) the polynucleotide sequences which hybridize under stringent conditions to the complementary sequences of (a); and

(c) polynucleotide sequences which are degenerate to polynucleotide sequences of (a) or (b).

4. A polynucleotide according to any one of the preceding claims, lacking the native leader sequences or any of the 5' or 3' untranslated regions of the polynucleotide.

5. A polynucleotide according to claim 4, wherein the native leader sequences or any of the 5' or 3' untranslated regions are replaced with exogenous control/regulatory sequences which regulate optimised/enhanced expression of the polynucleotide in an expression system.

6. A polynucleotide according to any one of the preceding claims which encodes codeinone reductase enzyme of *Papaver somniferum.*

7. A polynucleotide according to any one of claims 2 to 6, which is a synthetic polynucleotide comprising one or more codons preferred for expression in plant cells.

8. An isolated and purified polynucleotide which codes for prokaryotic or eukaryotic expression of a codeinone reductase enzyme from an alkaloid poppy plant, or a variant, analog or fragment thereof, wherein the polynucleotide is expressed in an environment selected from the group consisting of the extracellular environment, an intracellular membranous compartment, intracellular cytoplasmic compartment or combinations thereof.

9. A polynucleotide according to claim 8, comprising a nucleotide sequence which directs expression of the codeinone reductase enzyme with respect to a particular cellular compartment or the extracellular environment.

10. An isolated and purified polynucleotide having a sequence which is complementary to all or part of the sequence of a polynucleotide according to any one of claims 1 to 9.

11. A recombinant DNA construct comprising the polynucleotide according to any one of claims 1 to 10.

12. A DNA construct according to claim 11, which is a viral or plasmid vector.

13. A DNA construct according to claim 11 or claim 12, capable of directing prokaryotic or eukaryotic expression of the polynucleotide encoding a codeinone reductase enzyme.

14. A DNA construct according to any one of claims 11 to 13, comprising a promoter suitable to control the expression of the polynucleotide.

15. A DNA construct according to claim 14, wherein the promoter is endogenous.

16. A DNA construct according to claim 14, wherein the promoter is derived from nos, cauliflower mosaic virus or subterranean clover mosaic virus.

17. A DNA construct according to claim 12 wherein the plasmid is pCAL-c.

18. A DNA construct according to claim 12 wherein the plasmid is pGEM-T.

19. A DNA construct according to claim 12 wherein the plasmid is pFastBacI.

20. An isolated and purified codeinone reductase enzyme, being a product of prokaryotic or eukaryotic expression of the polynucleotide of any one of claims 1 to 10 or a DNA construct of any one of claims 11 to 19.

21. An enzyme according to claim 20, being a product of yeast cell expression.

22. An enzyme according to claim 20, being a product of bacterial cell expression.

23. An enzyme according to claim 20, being a product of animal cell expression.

24. An enzyme according to claim 23, being a product of insect cell expression.

25. An enzyme according to claim 20, being a product of plant cell expression.

26. An enzyme according to claim 25, wherein the plant cell is an alkaloid poppy plant cell.

27. An enzyme according to claim 26, wherein the alkaloid poppy is *Papaver somniferum*.

28. An enzyme according to any one of claims 20 to 27, which is a variant incorporating amino acid deletions, substitutions, additions or combinations thereof, wherein the variant retains one or more of the biological properties of codeinone reductase enzyme.

29. A cell transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19.

30. A cell according to claim 29, which is a plant cell.

31. A cell according to claim 30, wherein the plant cell is derived from an alkaloid poppy plant.

32. A cell according to claim 31, wherein the poppy plant is *Papaver somniferum*

33. A cell according to claim 29, which is a bacterial cell.

34. A cell according to claim 29, which is an animal cell.

35. A cell according to claim 29, which is a yeast cell.

36. A callus transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19.

37. A plant transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19, wherein the plant exhibits altered expression of the codeinone reductase enzyme.

38. A plant according to claim 37, wherein the altered expression is overexpression of the codeinone reductase enzyme.

39. A plant according to claim 37, wherein the altered expression is reduced expression of the codeinone reductase enzyme.

40. A plant according to any one of claims 37 to 39, which is an alkaloid poppy plant.

41. A plant according to claim 40, wherein the plant has a higher or different alkaloid content when compared to a plant which has not been so transformed or transfected.

42. A plant according to claim 40 or claim 41, wherein the alkaloid poppy plant is *Papaver somniferum.*

43. A method for preparing plants which overexpress a codeinone reductase enzyme, comprising transfecting or transforming a plant cell, a plant part or a plant, with the polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19.

44. A method according to claim 43, wherein the plant is an alkaloid poppy plant.

45. A method according to claim 44, wherein the poppy plant is *Papaver somniferum.*

46. A method of altering the yield or type of alkaloid in a plant comprising transforming or transfecting a plant cell, a plant part or a plant with a polynucleotide, or a variant, analog or fragment thereof, encoding a codeinone reductase enzyme or with a

polynucleotide which binds under stringent conditions to the polynucleotide encoding said enzyme.

47. A method of increasing the yield of alkaloid in a plant comprising transforming or transfecting a plant cell, a plant part or a plant with a polynucleotide, or a variant, analog or fragment thereof, encoding a codeinone reductase enzyme wherein the enzyme is overexpressed in said plant.

48. A method of altering type or blend of alkaloid in a plant comprising transforming or transfecting a plant cell, a plant part or a plant with a polynucleotide or a variant, analog or fragment thereof, encoding a codeinone reductase enzyme or with a polynucleotide which binds under stringent conditions to the polynucleotide encoding said enzyme.

49. A stand of stably reproducing alkaloid poppies transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19, having altered expression of the codeinone reductase enzyme.

50. A stand of stably reproducing alkaloid poppies transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19, having a higher or different alkaloid content when compared to a plant which has not been so transformed or transfected.

51. A stand of stably reproducing alkaloid poppies according to claim 49 or claim 50, wherein the alkaloid poppy is *Papaver somniferum.*

52. Straw of stably reproducing poppies according to any one of claims 49 to 51, having a higher or different alkaloid content when compared to the straw obtained from an alkaloid poppy which has not been transformed or transfected.

53. A concentrate of straw according to claim 52, having a higher or different alkaloid content when compared to the concentrate of straw obtained from an alkaloid poppy which has not been transformed or transfected.

54. An alkaloid when isolated from the straw according to any one of claims 49 to 52 or the concentrate according to claim 53.

55. An alkaloid according to claim 54, selected from the group consisting of morphine, codeine, oripavine and thebaine.

56. A method for the production of poppy plant alkaloids, comprising the steps of;

a) harvesting capsules of an alkaloid poppy plant transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according

to any one of claims 11 to 19, to produce a straw where the poppy plant is such a plant that the straw has a higher or different alkaloid content when compared to the straw obtained from a poppy plant which has not been transformed or transfected.

b) chemically extracting the alkaloids from the straw.

57. A method for the production of poppy alkaloids, comprising the steps of;

a) collecting and drying the latex of the immature capsules of an alkaloid poppy plant transformed or transfected with a polynucleotide according to any one of claims 1 to 10 or a DNA construct according to any one of claims 11 to 19, to produce opium wherein the poppy plant is such a plant that the opium has a higher or different alkaloid content when compared to the opium obtained from a poppy plant which has not been transformed or transfected.

b) chemically extracting the alkaloids from the opium.

58. A method according to claim 56 or claim 57, wherein the alkaloid is selected from the group consisting of morphine, codeine, oripavine and thebaine.

59. The polynucleotide sequence encoding codeinone reductase comprised in microbial deposit DSM 12737.

60. The polynucleotide sequence encoding codeinone reductase comprised in microbial deposit DSM 12738.

61. The polynucleotide sequence encoding codeinone reductase comprised in microbial deposit DSM 12739.

62. The polynucleotide sequence encoding codeinone reductase comprised in microbial deposit DSM 12740.

DATED this 26th day of March, 1999

JOHNSON & JOHNSON RESEARCH PTY LTD

Attorney: PAUL G. HARRISON
Fellow Institute of Patent Attorneys of Australia
of BALDWIN SHELSTON WATERS

Fig 1

SCHEME I

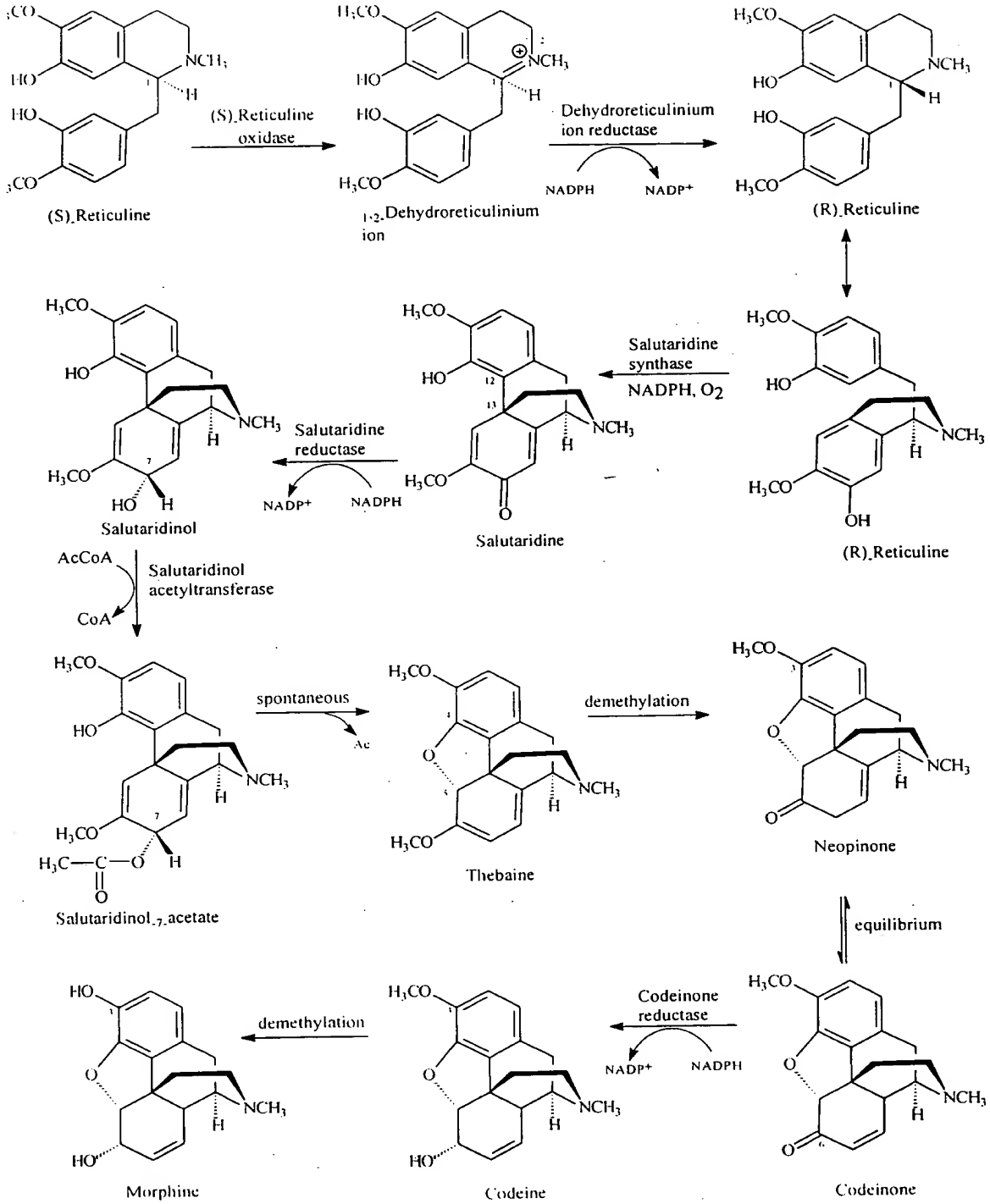


Fig 2

Peptide 3	X L Q E L M A
Peptide 7	V L H Q I A V A R G K
Peptide 14	D D D E L F I T S K
Peptide 16	I P D V V N Q V E M S P T L G Q
Peptide 17	X V N E I P K
Peptide 25	X V A Q V
Peptide 29	I F D N X L T A E D

Fig 3

```
            51                                                  90
Alfalfa     KQGYRHFDTA AAYGSEQALG EALKEAIELG LVTREELFVT
Glycyrrh.   KQGYRHFDTA AAYGSETALG EALKEARDLG LVTREELFVT
Soybean     KQGYRHFDTA AAYGSEQALG EALKEAIHLG LVSRQDLFVT
Opium poppy ..........  ..........  ..........  .....ELFIT

  91                                                        140
  SKLWVTENHP HLVIPALQKS LKTLQLDYLD LYLIHWPLSS QPGKFSFPID
  SKLWVTENHP HLVIPALRKS LETLQLEYLD LYLIHWPLSS QPGKFSFPIQ
  SKLWVTENHP HLVLPALRKS LKTLQLEYLD LYLIHWPLSS QPGKFSFPIE
  SK........  ..........  ..........  ..........  ..........

  141                                                       190
  VADLLPFDVK GVWESMEESL KLGLTKAIGV SNFSVKKLEN LLSVATVLPA
  VEDLLPFDVK GVWESMEECL KLGLTKAIGV SNFSVKKLQN LLSVATIRPA
  VEDLLPFDVK GVWESMEECQ KLGLTKAIGV SNFSVKKLQN LLSVATIRPV
  ..........  ..........  ..........  .......LQE LMA...IPDV

  191                                                       240
  VNQVEMN...  LAWQQKKLRE FCNANGIVLT AFSPLRKGAS RGPNEVMEND
  VNQVEMN...  LAWQQKKLRE FCTANGIVLT AFSPLRKGAS RGPNEVMEND
  VDQVEMN...  LAWQQKKLRE FCKENGIIVT AFSPLRKGAS RGPNEVMEND
  VNQVEMSPTL..........  ..........  ..........  ..........

  241
  MLKEIADAHG KSVAQISLRW LYEQGVTFVP KSYDKERMNQ NLC
  MLKGIAEAHG KSIAQVSLRW LYEQGVTFVA KSYDKERMNQ NLQ
  VLKEIAEAHG KSIAQVSLRW LYEQGVTFVP KSYDKERMNQ NLH
  VLHQIAVARG K.........  .....VNEIP K.........  ...
```

Fig 4

```
cor1.1   .MESNGVPMI TLSSG...IR MPALGMGTAE TMVKGTEREK LAFLKAIEVG
cor1.2   .--------- ---------- --------V- --E------- ----N-----
cor1.3   .--------- ---------- ---------- ---------- ----------
cor1.4   .--------- ---------- ---------- ---------- ----------
6'dcs    MAAAIEI-T- VFPNSSAQQ- --VV---S-P DFTCKKDT.- E-IIE-VKQ-

cor1.1   YRHFDTAAAY QTEECLGEAI AEALQLGLIK SRDELFITSK LWCADAHADL
cor1.2   ---------- -S-------- ---------- ---------- ----------
cor1.3   ---------- -S-------- ---------- ---------- ----------
cor1.4   ---------- -S-------- ---------- ---------- ----------
6'dcs    ---------- GS-QA----L K--IH---VS .-QD-V---- --VTEN-PH-

cor1.1   VLPALQNSLR NLKLDYLDLY LIHHPVSLKP GKFVNEIPKD HILPMDYKSV
cor1.2   ---------- ----E----- ---------- --L------- ----------
cor1.3   ---------- ---------- ---------- ---------- ----------
cor1.4   ---------- ----E----- ---------- ---------- ----------
6'dcs    -----RK--K T-Q-E----- ---W-L-SQ- ---SFP-EVE DL--F-V-G-

cor1.1   WAAMEECQTL GFTRAIGVCN FSCKRLQELM ETANSPPVVN QVEMSPTLHQ
cor1.2   ---------- --------S- ----K----- A--KI----- ----------
cor1.3   ---------- ---------- ----K----- AA-KI----- ----------
cor1.4   ---------- --------S- ----K----- AA-KI----- ----------
6'dcs    -ES-----K- -L-K----S- --V-K--N-L SV-TIR---D ----NLAWQ-

cor1.1   KNLREYCKAN NIMITAHSVL GAVGAAWGTN AVMHSKVLHQ IAVARGKSVA
cor1.2   ---------- ---------- --I--P--S- ---D------ ----------
cor1.3   ---------- ---------- --IC-P--S- ---D------ ----------
cor1.4   ---------- ---------- --I--P--S- ---D------ ----------
6'dcs    -K---F--E- G-IV--F-P- .RK--SR-P- E--END--KE --E-H---I-

cor1.1   QVSMRWVYQQ GASLVVKSFN EARMKENLKI FDWELTAEDM EKISEIPQSR
cor1.2   ---------- ---------- ---------- --S------- ----------
cor1.3   ---------- ---------- -G-------- --------N- ----------
cor1.4   ---------- ---------- -G-------- ---------- ----------
6'dcs    ---L--L-E- -VTF-P--YD KE--NQ--H- ---A--EQ-H H---Q-S---

cor1.1   TSSAAFLLSP TGPFKTEEEF WDEKD
cor1.2   ----D----- ---------- -----
cor1.3   ----D----- ---------- -----
cor1.4   ---------- ---------- -----
6'dcs    .....LISG- -K-..QLADL --DQI
```

FIGURE 5

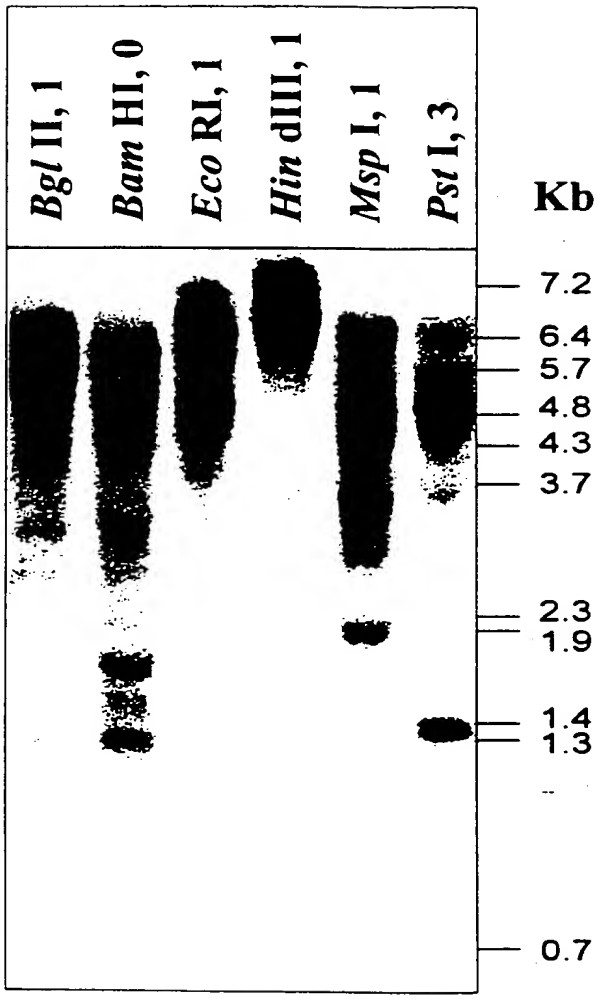

Bgl II, 1
Bam HI, 0
Eco RI, 1
Hin dIII, 1
Msp I, 1
Pst I, 3
Kb
7.2
6.4
5.7
4.8
4.3
3.7
2.3
1.9
1.4
1.3
0.7

FIGURE 6

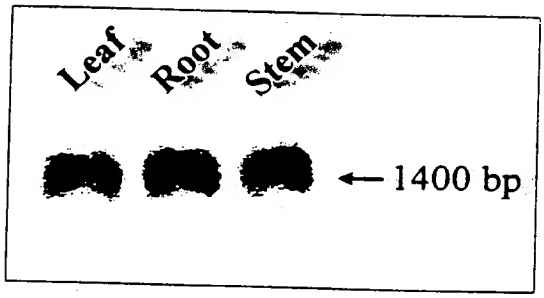


FIGURE 7

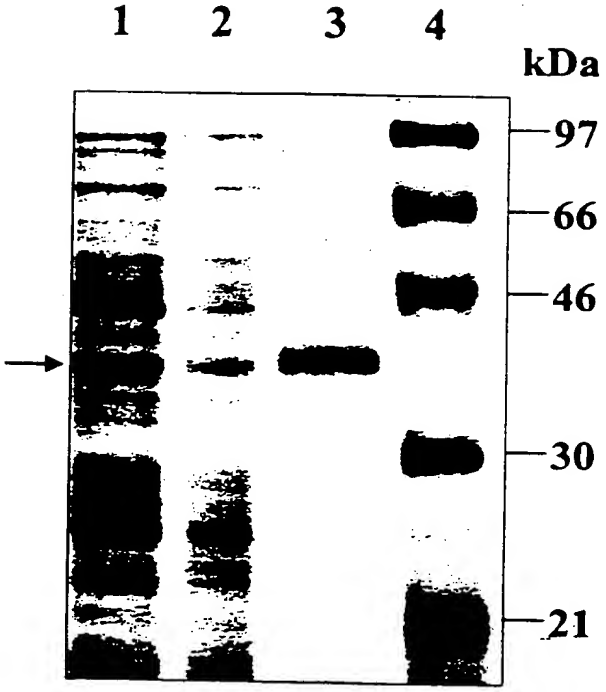
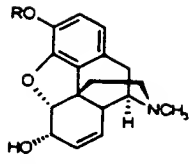
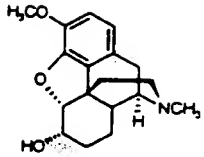


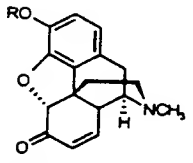
FIGURE 8



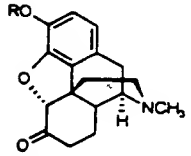
Substrate	R	K_m
Codeine	CH_3	187 µM
Morphine	H	200 µM



Substrate	K_m
Dihydrocodeine	628 µM

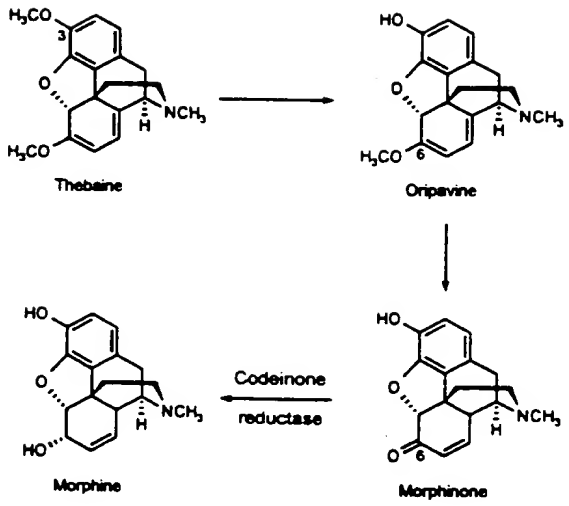


Substrate	R	K_m
Codeinone	CH_3	48 µM
Morphinone	H	143 µM



Substrate	R	K_m
Hydrocodone	CH_3	55 µM
Hydromorphone	H	90 µM

FIGURE 9


Thebaine
Oripavine
Morphinone
Morphine
Codeinone
reductase

cor1.1
cds6-971

Fig 10

GAAAAATGGAGAGTAATGGTGTACCTATGATCACTCTCAGTTCCGGCATTCGGATGCCTGCTTTAGGTATGGGAA
CAGCTGAAACAATGGTAAAAGGAACAGAAAGAGAGAAATTGGCGTTTTTGAAAGCGATAGAGGTCGGTTACAGAC
ACTTCGATACAGCTGCTGCATACCAAACTGAAGAGTGTCTTGGTGAAGCTATAGCTGAAGCACTTCAACTTGGTC
TAATAAAATCTCGAGATGAACTCTTCATCACTTCCAAGCTCTGGTGCGCTGATGCTCACGCTGATCTTGTCCTCC
CTGCTCTTCAGAATTCTCTGAGGAATCTTAAATTGGACTATCTTGATCTATATTTGATACACCATCCGGTAAGCT
TGAAGCCAGGGAAGTTTGTTAACGAAATACCAAAGGATCATATCCTTCCAATGGACTACAAATCTGTATGGGCAG
CCATGGAAGAGTGTCAGACCCTTGGCTTCACTAGGGCAATCGGGGTCTGTAATTTCTCATGCAAAAGGCTTCAAG
AGTTGATGGAAACAGCCAACAGCCCTCCAGTTGTGAATCAAGTGGAGATGAGCCCGACTTTACATCAAAAAAATC
TGAGGGAATATTGCAAGGCCAATAATATCATGATCACCGCACACTCAGTTTTGGGAGCCGTAGGTGCCGCCTGGG
GCACCAATGCAGTTATGCATTCTAAGGTGCTTCACCAGATTGCTGTGGCCAGAGGAAAATCTGTTGCCCAGGTTA
GTATGAGATGGGTTTACCAGCAAGGCGCGAGTCTTGTGGTGAAAAGTTTCAATGAAGCGAGGATGAAGGAAAACC
TTAAGATATTTGATTGGGAACTAACGGCAGAAGACATGGAAAAGATCAGTGAGATTCCACAATCTAGAACAAGCT
CTGCTGCTTTCTTGTTATCACCGACTGGACCTTTCAAAACTGAAGAAGAGTTCTGGGATGAGAAGGATTGAAACA
TCAATTATAGATGGTAAGTGAGGACTGTCAAAAAAGTAATCAGTTTTTCCCTCCGTTTTG

cor1.2
cds 1-966

Fig 11

ATGGAGAGTAATGGTGTACCTATGATCACTCTCAGTTCCGGCATTCGGATGCCTGCTTTAGGTATGGGAACAGTT
GAAACAATGGAAAAGGGAACAGAAAGAGAGAAATTGGCGTTTTTGAATGCGATAGAGGTCGGTTACAGACACTTC
GATACAGCTGCTGCATACCAAAGTGAAGAGTGTCTTGGTGAAGCTATAGCTGAAGCACTTCAACTTGGTTTAATA
AAATCTCGAGATGAACTCTTCATCACTTCCAAGCTCTGGTGCGCTGATGCTCACGCTGATCTTGTCCTCCCTGCT
CTTCAGAATTCTCTGAGGAATCTCAAATTGGAGTACCTTGATCTATATTTGATACACCATCCGGTAAGCTTGAAG
CCAGGGAAGCTTGTTAACGAAATACCAAAGGATCATATTCTTCCAATGGACTACAAATCTGTATGGGCAGCCATG
GAAGAGTGTCAGACCCTTGGCTTCACTAGGGCAATCGGTGTCAGTAATTTCTCATGCAAAAAGCTTCAAGAGTTG
ATGGCAACAGCCAAGATCCCTCCAGTTGTGAATCAAGTGGAGATGAGCCCGACTTTACATCAAAAAAATCTGAGG
GAATATTGCAAGGCCAATAATATCATGATCACTGCACACTCGGTTTTGGGAGCCATAGGTGCTCCATGGGGCAGC
AACGCAGTTATGGATTCTAAGGTGCTTCACCAGATTGCTGTGGCAAGAGGAAAATCTGTTGCCCAGGTTAGTATG
AGATGGGTTTACCAGCAAGGCGCGAGTCTTGTGGTGAAAAGTTTCAATGAAGCGAGGATGAAGGAAAACCTTAAG
ATATTTGATTCGGAACTAACGGCAGAAGATATGGAAAAGATCAGTGAGATTCCGCAATCTAGAACAAGCTCTGCT
GATTTCTTGTTATCACCGACTGGACCTTTCAAAACTGAAGAAGAGTTCTGGGATGAGAAGGATTGA

cor1.3
cds1-966

Fig 12

ATGGAGAGTAATGGTGTACCTATGATCACTCTCAGTTCCGGCATTCGGATGCCTGCTTTAGGTATGGGAACAGCT
GAAACAATGGTAAAAGGAACAGAAAGAGAGAAATTGGCGTTTTTGAAAGCGATAGAGGTCGGTTACAGACACTTC
GATACAGCTGCTGCATACCAAAGTGAAGAGTGTCTTGGTGAAGCTATAGCTGAAGCACTTCAACTTGGTCTAATA
AAATCTCGAGATGAACTCTTCATCACTTCCAAGCTCTGGTGCGCTGATGCTCACGCTGATCTTGTCCTCCCTGCT
CTTCAGAATTCTCTGAGGAATCTTAAATTGGACTATCTTGATCTATATTTGATACACCATCCGGTAAGCTTGAAG
CCAGGGAAGTTTGTTAACGAAATACCAAAGGATCATATCCTTCCAATGGACTACAAATCTGTATGGGCAGCCATG
GAAGAGTGTCAGACCCTTGGCTTCACTAGGGCAATCGGGGTCTGTAATTTCTCATGCAAAAAGCTTCAAGAGTTG
ATGGCAGCAGCCAAGATCCCTCCAGTTGTGAATCAAGTGGAGATGAGCCCGACTTTACATCAAAAAAATCTGAGG
GAATATTGCAAGGCCAATAATATCATGATCACTGCACACTCGGTTTTGGGAGCCATATGTGCTCCATGGGGCAGC
AATGCAGTTATGGATTCTAAGGTGCTTCACCAGATTGCTGTGGCAAGAGGAAAATCTGTTGCCCAGGTTAGTATG
AGATGGGTTTACCAGCAAGGCGCGAGTCTAGTGGTGAAAAGTTTCAATGAAGGGAGGATGAAGGAAAACCTTAAG
ATATTTGATTGGGAACTAACGGCAGAGAATATGGAAAAGATCAGTGAGATTCCGCAATCTAGAACAAGCTCTGCT
GATTTCTTGTTATCACCGACTGGACCTTTCAAAACTGAAGAAGAGTTCTGGGATGAGAAGGATTGA

cor1.4
cds1-966

Fig 13

ATGGAGAGTAATGGTGTACCTATGATCACTCTCAGTTCCGGCATTCGGATGCCTGCTTTAGGTATGGGAACAGCT
GAAACAATGGTAAAAGGAACAGAAAGAGAGAAATTGGCGTTTTTGAAAGCGATAGAGGTCGGTTACAGACACTTC
GATACAGCTGCTGCATACCAAAGTGAAGAGTGTCTTGGTGAAGCTATAGCTGAAGCACTTCAACTTGGTTTAATA
AAATCTCGAGATGAACTCTTCATCACTTCCAAGCTCTGGTGCGCTGATGCTCACGCTGATCTTGTCCTCCCTGCT

CTTCAGAATTCTCTGAGGAATCTCAAATTGGAGTATCTTGATCTATATTTGATACACCATCCGGTAAGCTTGAAG
CCAGGGAAATTTGTTAACGAAATACCAAAGGATCATATTCTTCCAATGGACTACAAATCTGTATGGGCAGCCATG
GAAGAGTGTCAGACCCTTGGCTTCACTAGGGCAATCGGTGTCAGTAATTTCTCATGCAAAAAGCTTCAAGAGTTG
ATGGCAGCAGCCAAGATCCCTCCAGTTGTGAATCAAGTGGAGATGAGCCCTACTTTACATCAAAAAAATCTGAGG
GAATATTGCAAGGCCAATAATATCATGATCACTGCACACTCGGTTTTGGGAGCCATAGGTGCTCCATGGGGCAGC
AATGCAGTTATGGATTCTAAGGTGCTTCACCAGATTGCTGTGGCAAGAGGAAAATCTGTTGCCCAGGTTAGTATG
AGATGGGTTTACCAGCAAGGCGCGAGTCTTGTGGTGAAAAGTTTCAATGAAGGGAGGATGAAGGAAAACCTTAAG
ATATTTGATTGGGAACTAACGGCAGAAGATATGGAAAAGATCAGTGAGATTCCGCAATCTAGAACAAGCTCTGCT
GCTTTCTTGTTATCACCGACTGGACCTTTCAAAACTGAAGAAGAGTTCTGGGATGAGAAGGATTGA

cor1.5
partial seq

Fig 14

TGTGGTGAATCAGGTGGAGATGTGGCCGACTTTACATCAAAAAAATCTGAGGGAATATTGCAAGGCCAATAATAT
CATGATCACTGCACACTCGGTTTTGGGAGCCATAGGTGCTCCATGGGGCAGCAATGCAGTTATGGATTCTAAGGT
GCTT

cor1.6
partial seq

Fig 15

CTCTGGTGCGCTGATGCTCACGCTGATCTTGTCCTCCCTGCTCTTCAGAATTCTCTGAGGAATCTCAAATTGGAC
TACCTTGATCTATATTTGATACACCATCCGGTAAGCTTGAAGCCAGGGAAGCTTGTTAACGAAATACCAAAGGAT
CATATTCTTCCAATGGACTACAAATCTGTATGGGCAGCCATGGAAGAGTGTCAGACCCTTGGCTTCACTAGGGCA
ATCGGTGTCAGTAATTTCTCATGCAAAAAGCTTCAAGAGTTGATGGCAACAGCCAAGATCCCTCCA